

Toward Atomic Resolution CryoEM with Bioinformatics

Hong Zhou, Ph.D.

University of Texas - Medical School at Houston

Pathology & Laboratory Medicine

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Outlines

- **Introduction of our model system dsRNA virus and cytoplasmic polyhedrosis virus (CPV)**
- **High resolution imaging of CPV**
- **Data processing, software development**
- **CPV structures at 5 Å resolutions**
- **Atomic modeling by integrative bioinformatics approach**

Push the limit: dsRNA viruses as models

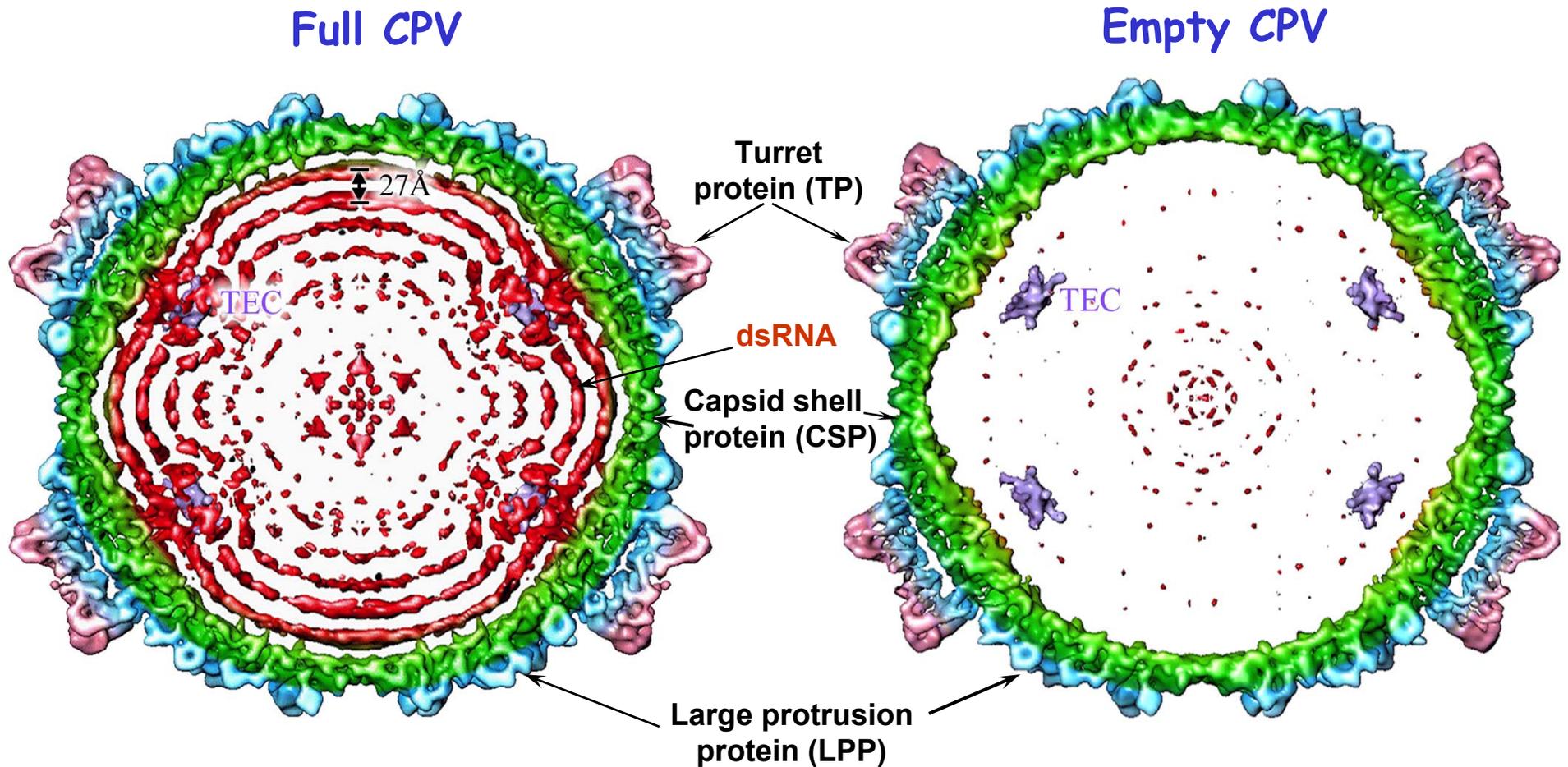
Genus	Host ranges	3D structure	# RNA segments	# Structural Proteins	Capsid size (Å)
Orthoreovirus	Mammals	X-ray & cryoEM	10	8	850
Rotavirus	Mammals/birds	cryoEM	11	6	800/1050
Orbivirus (BTV)	Mammals/insect vectors	X-ray & cryoEM	10	7	800
Phytoreovirus (rice dwarf virus)	Plants/insect vectors	cryoEM & X-ray	12	12	780
Cypovirus (CPV)	Insects	cryoEM	10	5	590/800
Fijivirus	Plant/insect vectors	no	10	11	700
Coltivirus	Mammals,invertebrate	no	12	12	800
Aquareovirus	Bony fish, crustaceans	cryoEM	11	7	800
Seadornavirus	Mammals	no	12	12	800
Mycoreovirus	fungus	no	11	10(?)	800(?)
Oryzavirus	Plant,invertebrate	no	10	10	700

Capable of endogenous mRNA transcription, capping & efficient release

Facts of CPV

- **Single-shelled capsid, yet very STABLE**
- **Fully capable of endogenous transcription, mRNA capping and release within intact virus**
- **Used as a bio-control agent, an environment-friendly pesticide**

Structural Organization of the CPV (13 Å)



TEC: Transcriptional Enzyme Complex

CryoEM imaging of CPV

- Liquid helium-cooled specimen (4 K) (JEOL)
- Gold aperture
- Fully "Baked" Quantifoil holey grids
- 300kV, field emission gun (FEG)
- Kodak SO163 films at 60,000 ×
- Focal pair, combined dose about 50 $e^-/\text{Å}^2$
- First micrograph < 1 μm defocus to improve accuracy of CTF correction
- Second micrograph, crucial, but only used in initial processing stage

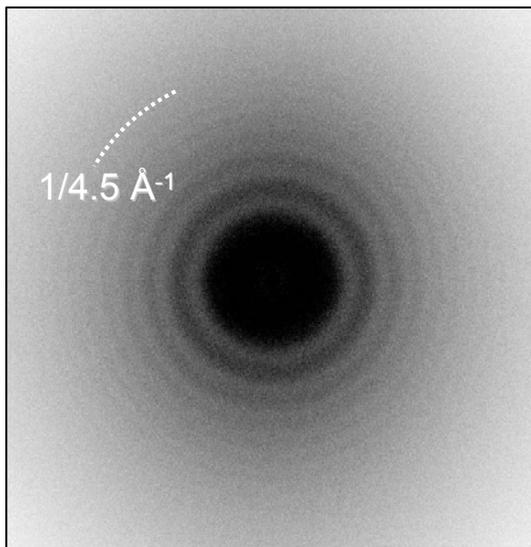
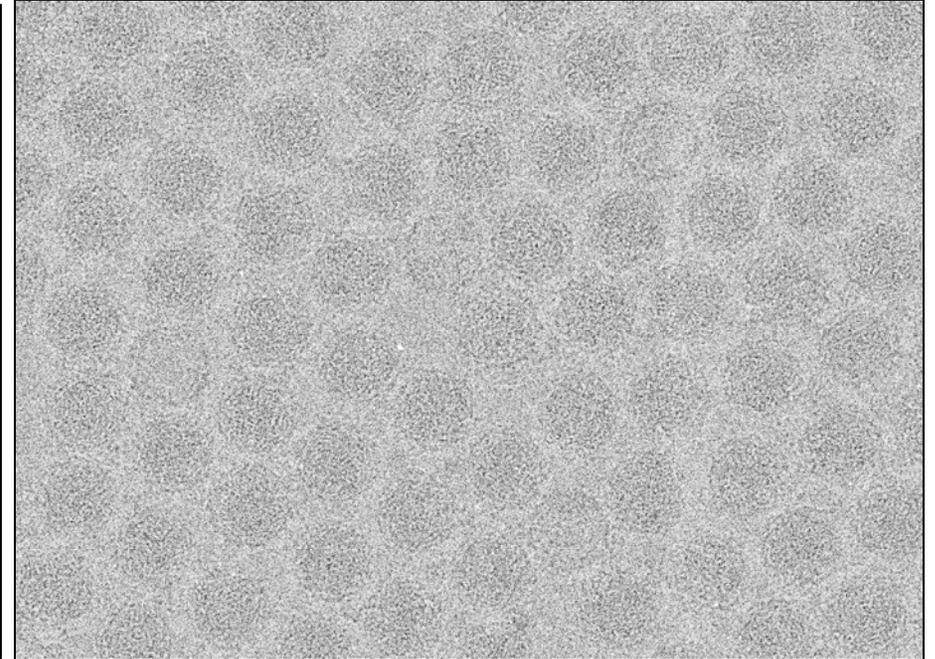
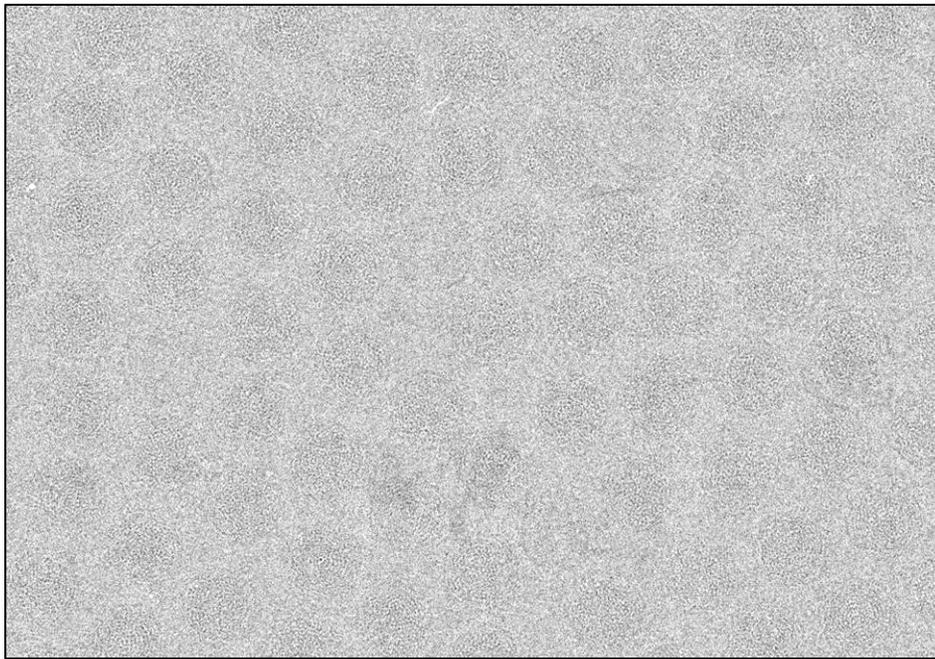


JEOL FEG 300 kV, liquid helium-cooled
National Center for Macromolecular Imaging,
Baylor College of Medicine

400kV JEOL4000

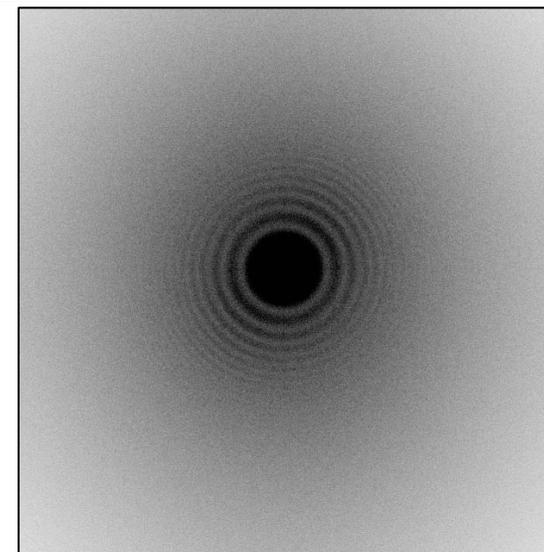
300kV FEI Polara G² F30 liquid helium (UT)

Image evaluation: incoherent averaging of Fourier transforms



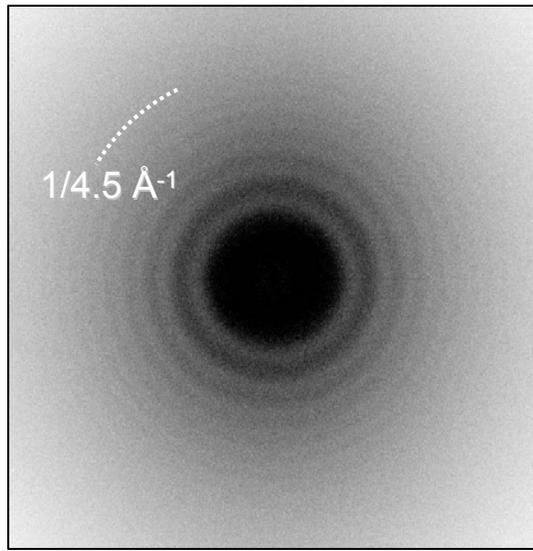
First micrograph

Incoherent average
of particle Fourier
transforms

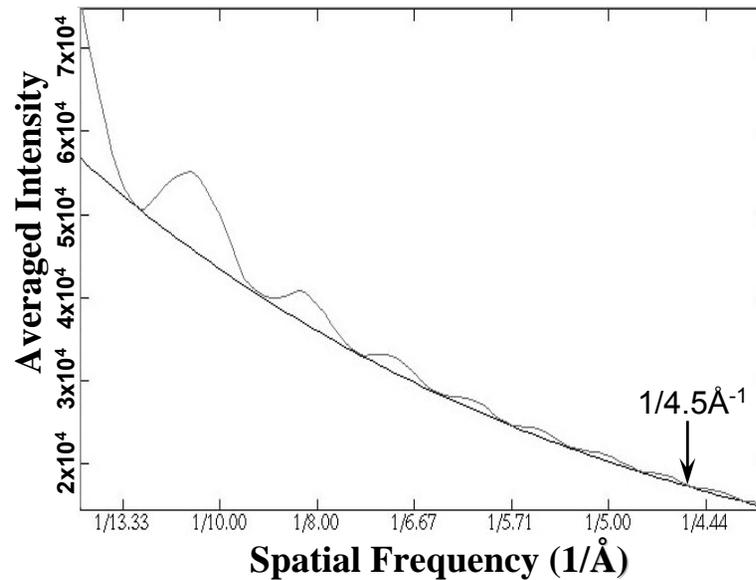


Second micrograph

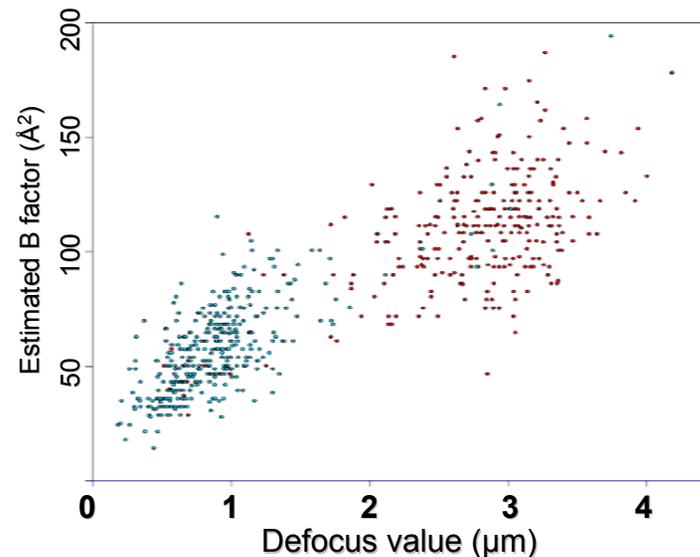
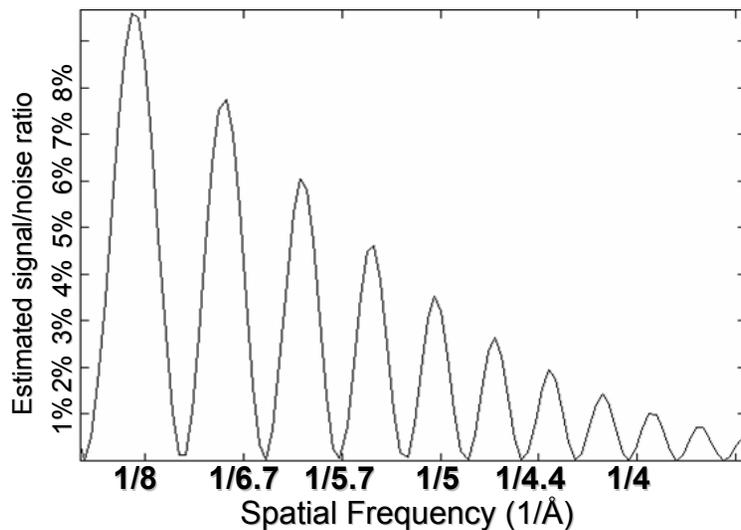
Defocus and "B" factor: exponential decay of data



Incoherent average of FTs



➤ Tens or even hundreds of thousands of particle images might be needed toward 4-5 Å resolution



IMIRS: an integrative and modular approach

- Integrated Management & Icos. Reconst. System

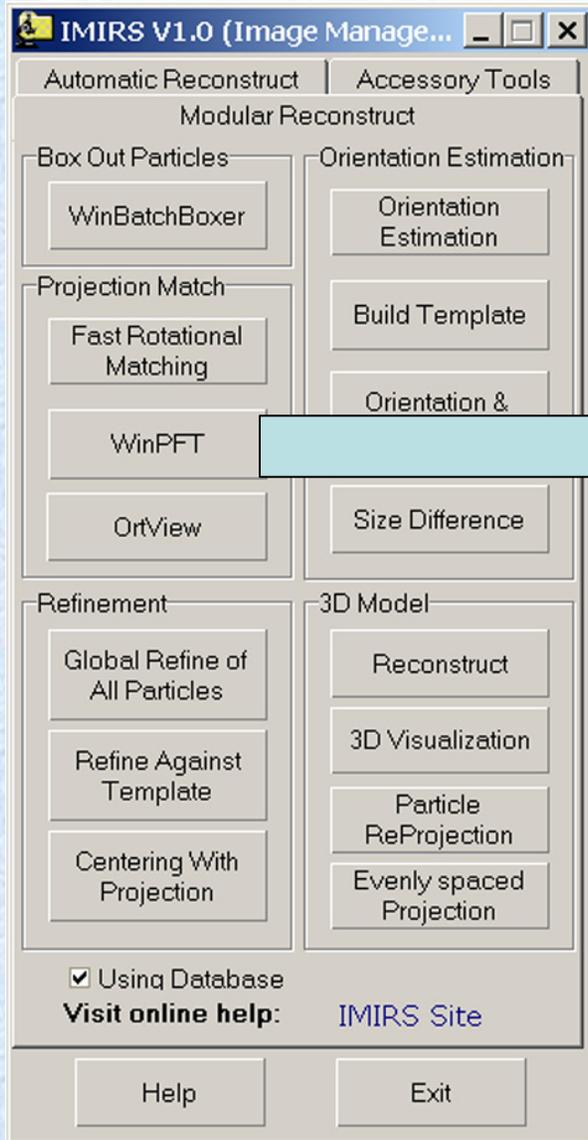
The image displays a central 3D model of a virus particle, surrounded by several screenshots of the IMIRS software interface. The screenshots show various modules and settings, including:

- winPFT V2.01**: A window for processing micrographs, showing options for 'User Type' (Beginning User or Experienced User), 'Please Select Process' (Cut Mic Images by 1 Pixel, Convert MRC to PIF), and 'Imaging Info' (Pixel size, Accelerating Voltage, Amplitude Contrast Ratio, Astigmatism Angle).
- IMIRS V1.0 (Image Management & Icosahedral Reconstruction System)**: The main interface, showing 'Modular Reconstruct' options (Automatic Reconstruct, Modular Reconstruct) and 'Process Tools' (Little Ending TO Big Ending, WinBoxer, TIFF -> MRC, Generate CTF, Project3File, maskTrans).
- IMIRS V1.0 Tools**: A window with a 'Gauge' and 'Image Size' settings (Image Size: 140, Display Circle: 70).
- rselectPointsList**: A window showing 'Out of Focus List' and 'In-Focus List' with coordinates (x, y).
- 3D Model**: A window showing a 'Refinement' and '3D Model' section, with a '3D Visualization' option.
- IMIRS V1.0 (Image Management & Icosahedral Reconstruction System) Running Panel**: A window showing a table of micrograph data.

MicrographID	ProjectID	ExperimentID	MicrographName
1	1	1	1767
2	1	1	1769
3	1	1	1771
4	1	1	1670
7	1	1	1657

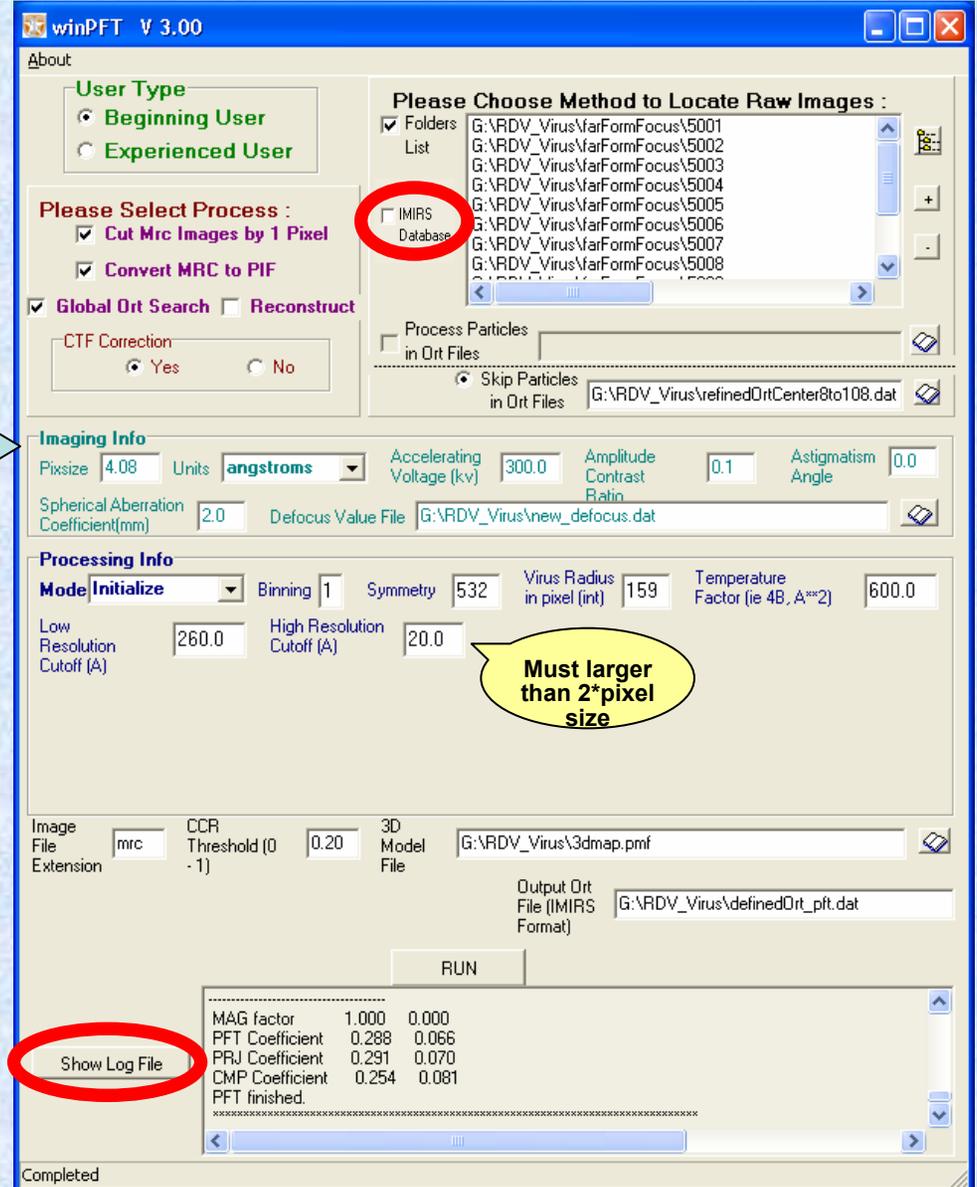
Engineering efforts of IMIRS

- Partially compatible with Pam's rules



PFT by Baker *et al.*

Demo



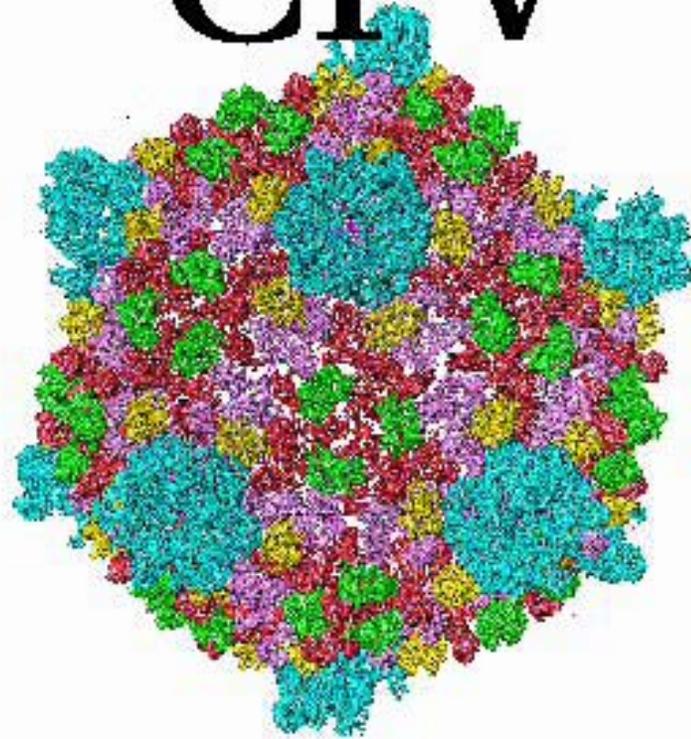
Summary of data processing statistics

- Number of focal pairs scanned: >1,000 pairs
- Number of focal pairs refined: 646 pairs
- 1.16 Å/pixel, 800x800 particle
- Number of particles processed: 135,000

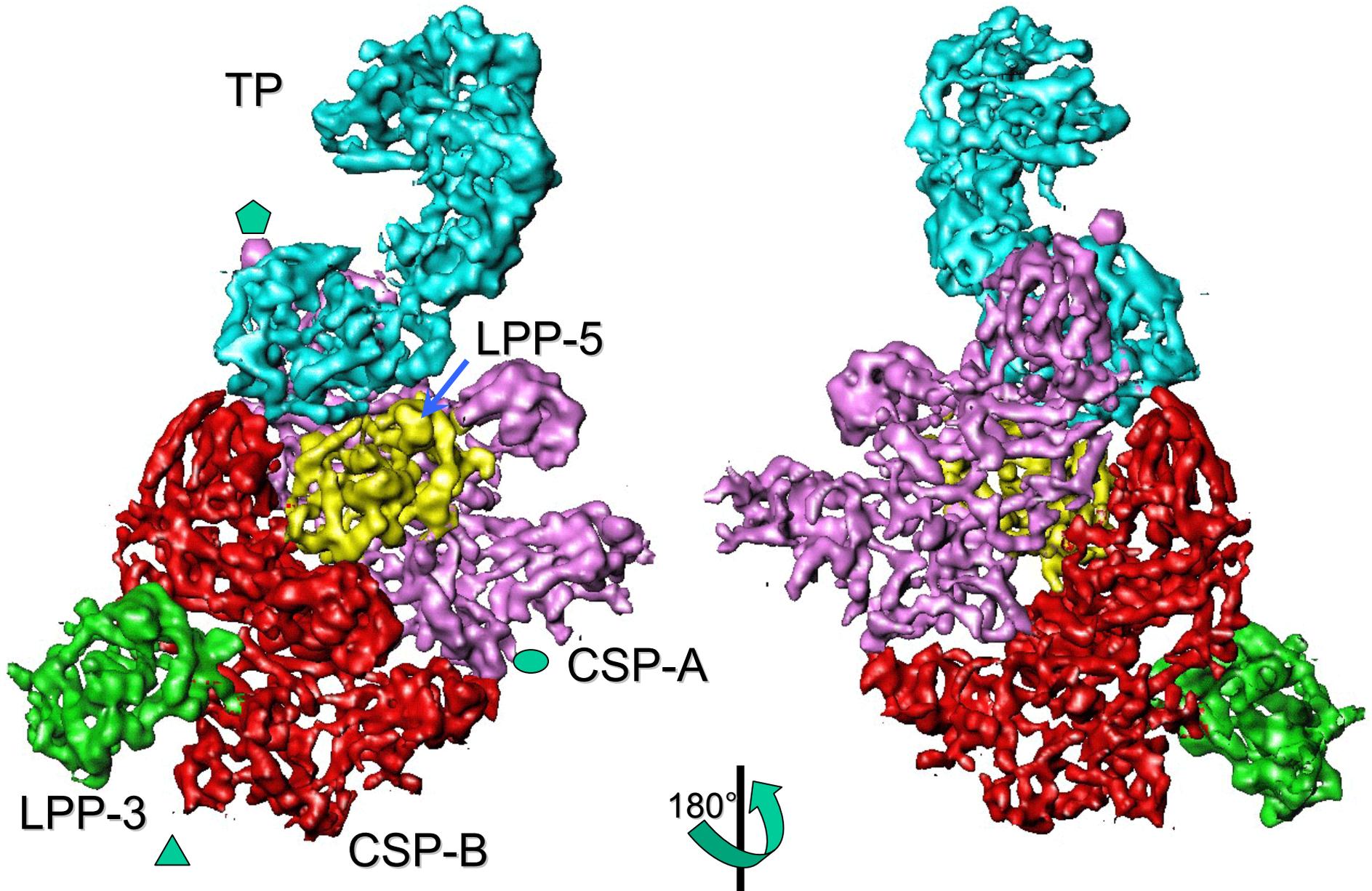
- Defocus ranges:
 - 1.9-3.7 μm and 0.2-1.7 μm for 1st & 2nd micrographs
- B factor: 100-210 and 40-140 Å² respectively
- Final reconstruction
 - 25,705** particle images used, all close-to-focus
 - refined to 1/3.5 Å⁻¹
 - effective resolution 5.2 Å
- Total averaging is about 1.5 million (25,705 x 60)

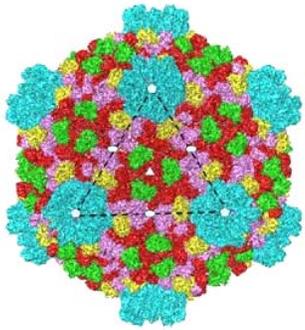
CPV Capsid Shell at 5.5 Å

CPV

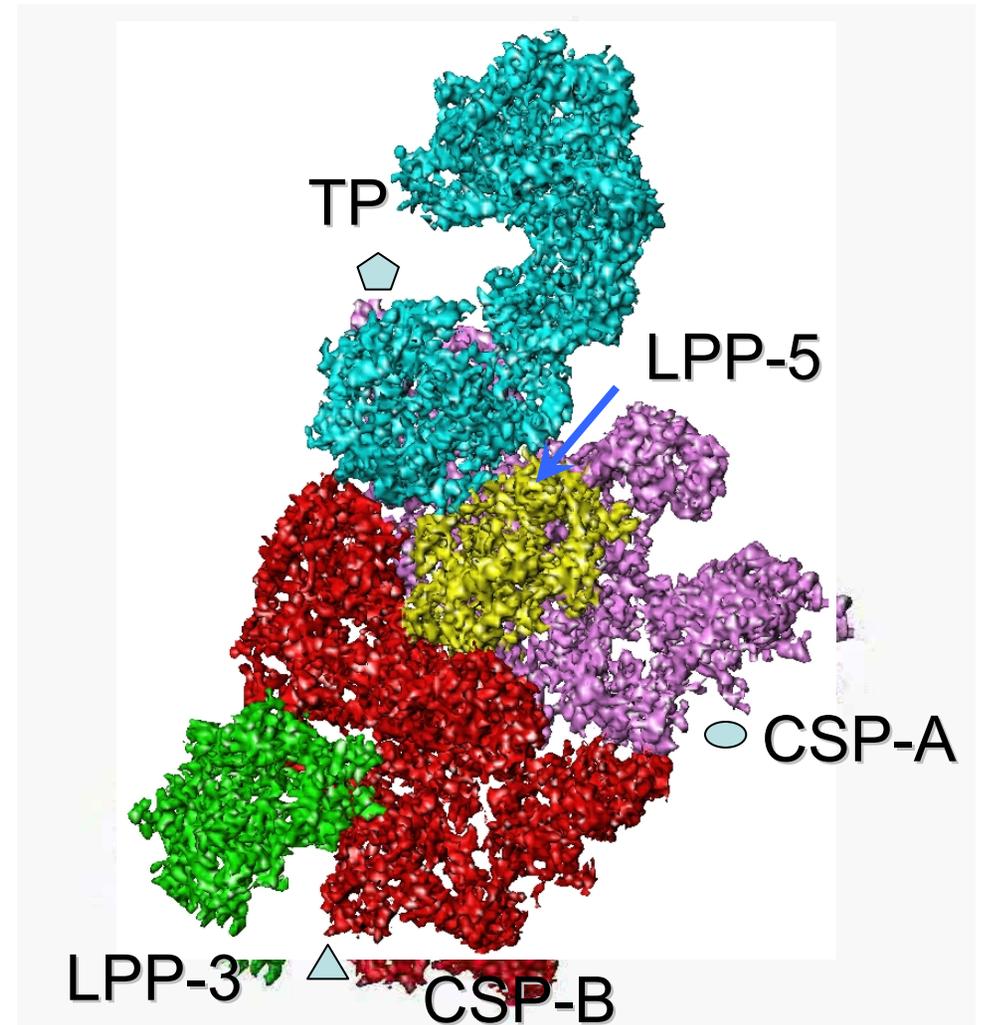
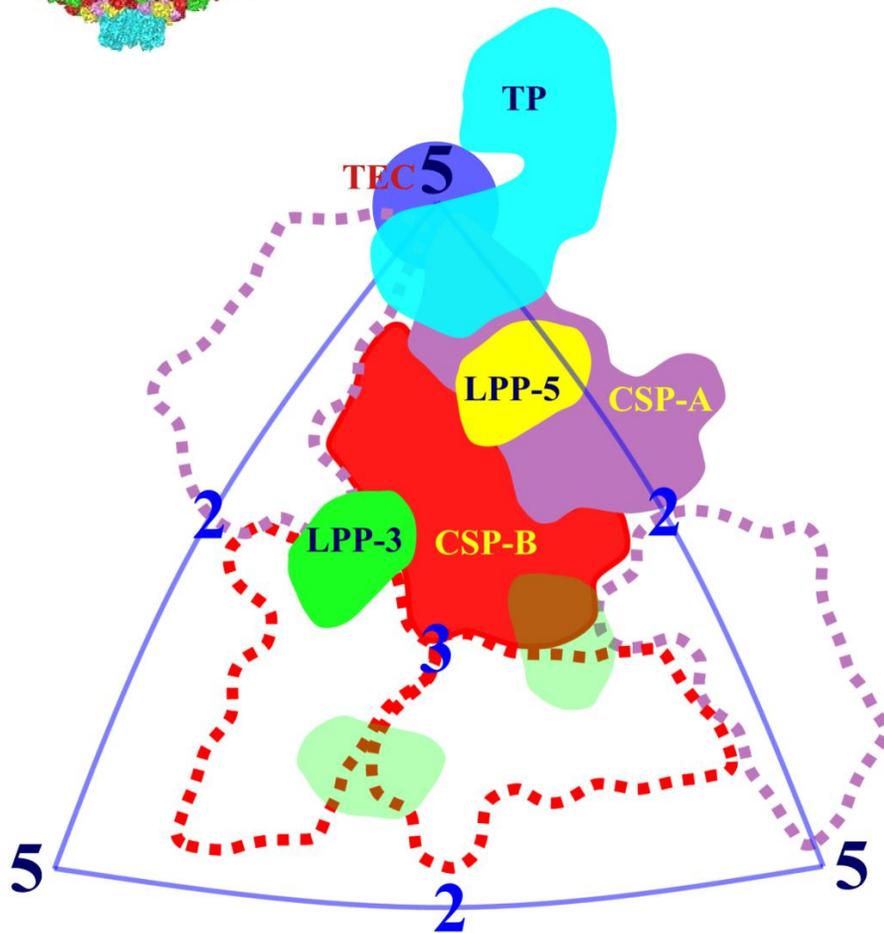


Asymmetric Unit: Molecular Interactions



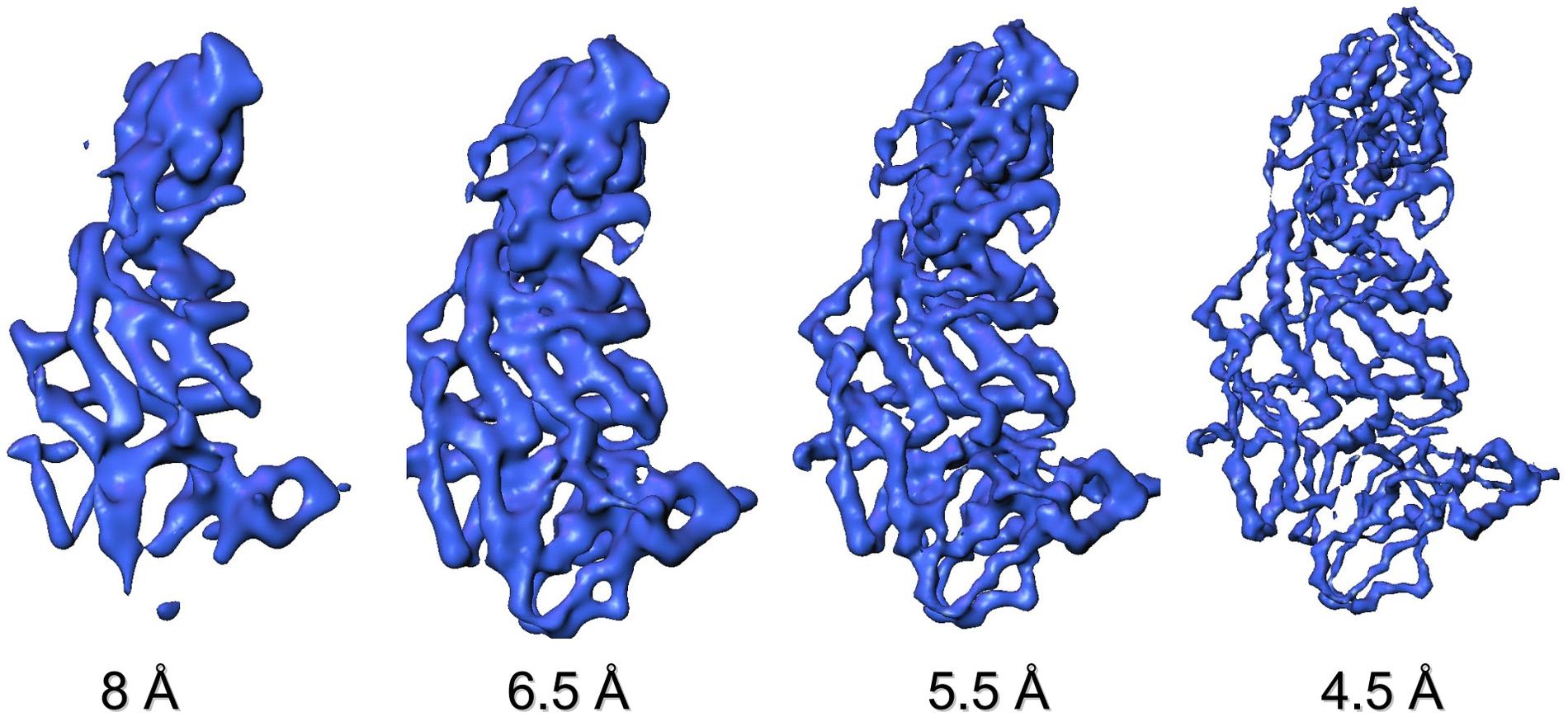


Asymmetric Unit of CPV

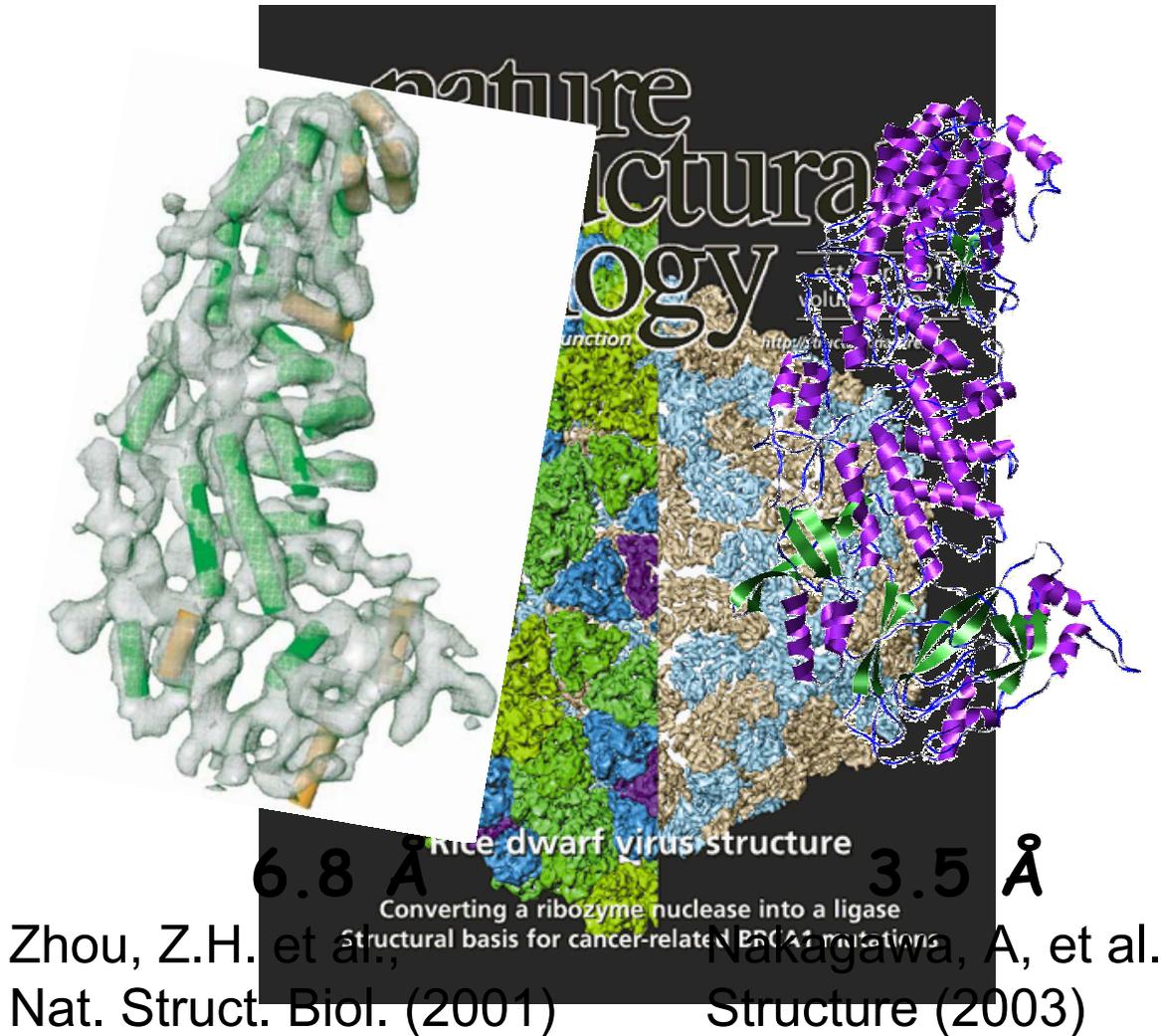


- Intensive molecular interactions
- Molecular clamps

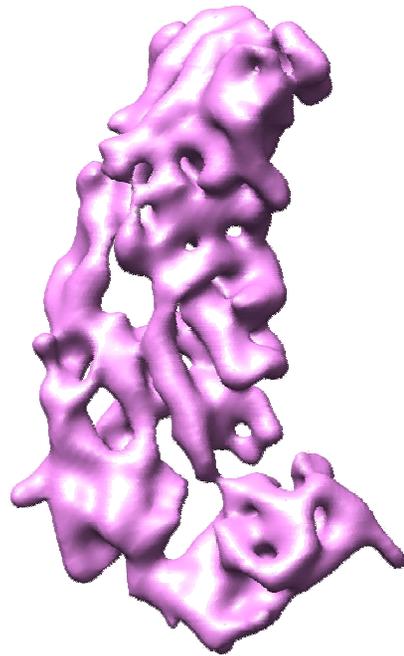
BTV CSP density map at different resolutions



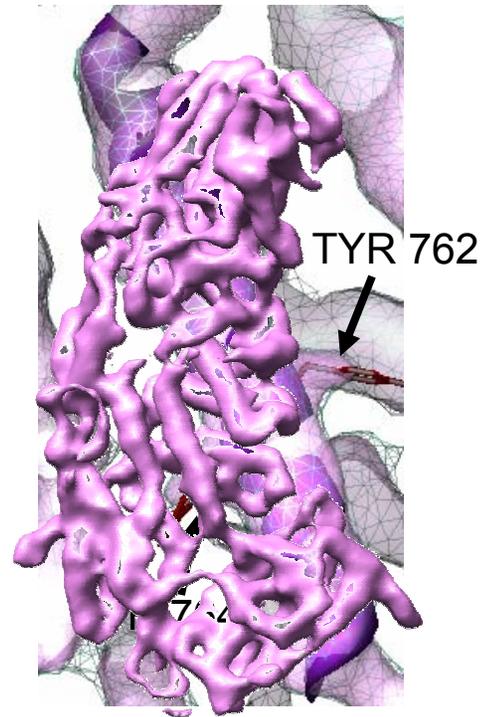
Simulation: What Can We See at 5-Å? (RDV: Rice Dwarf Virus)



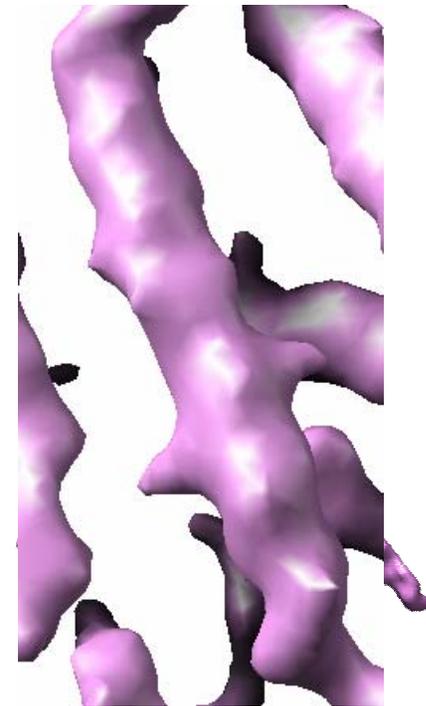
Bulky Side Chains Can Be Resolved at 5Å



8 Å



6 Å



5 Å

Bulky side chains of TYR, TRP, PHE, etc can be resolved at 5 Å

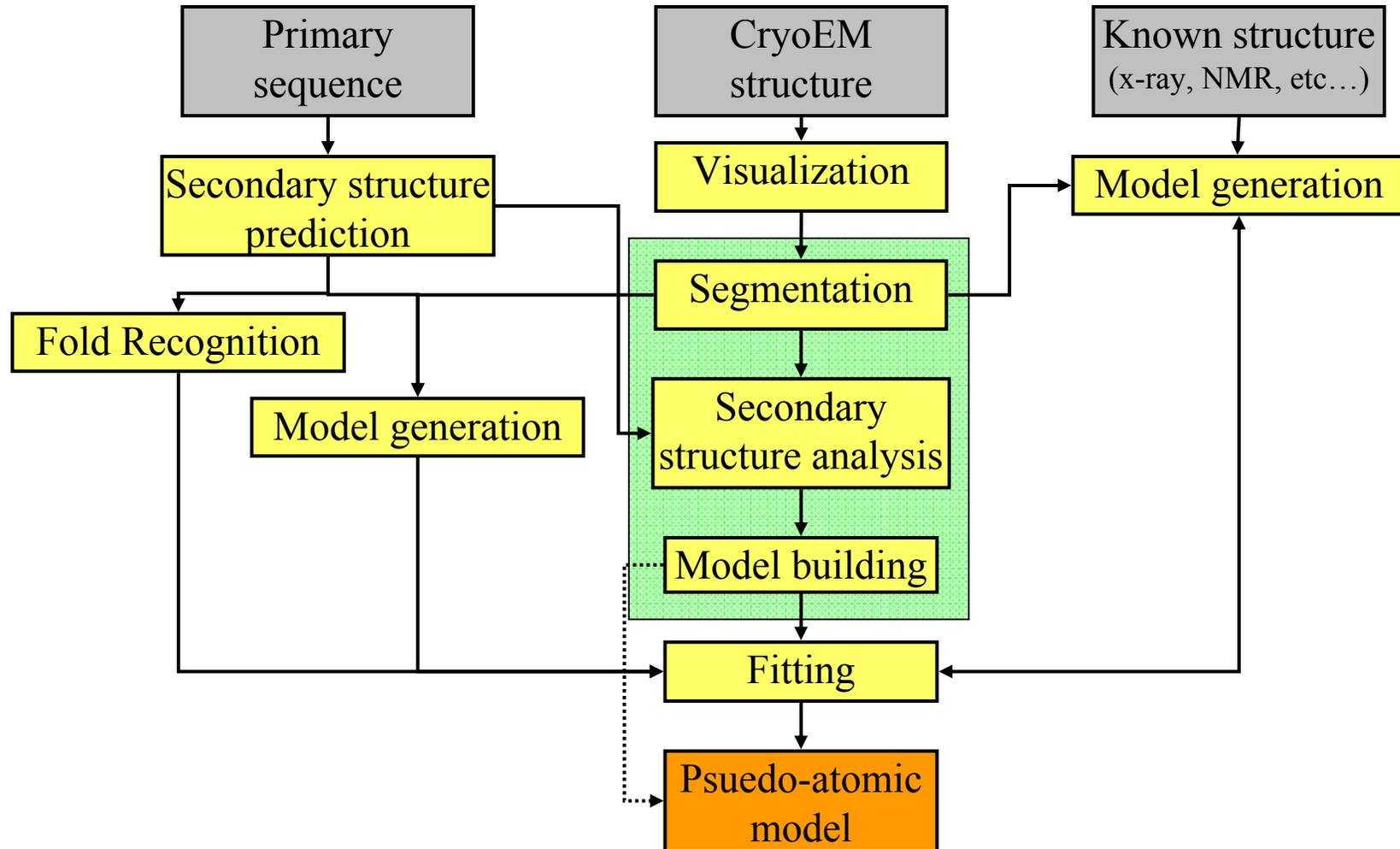
Approaches to Atomic Modeling of CryoEM Structures

Motivation: bottom-up approach (O, MAID, X-Build etc) NOT applicable to near-atomic resolution cryoEM maps

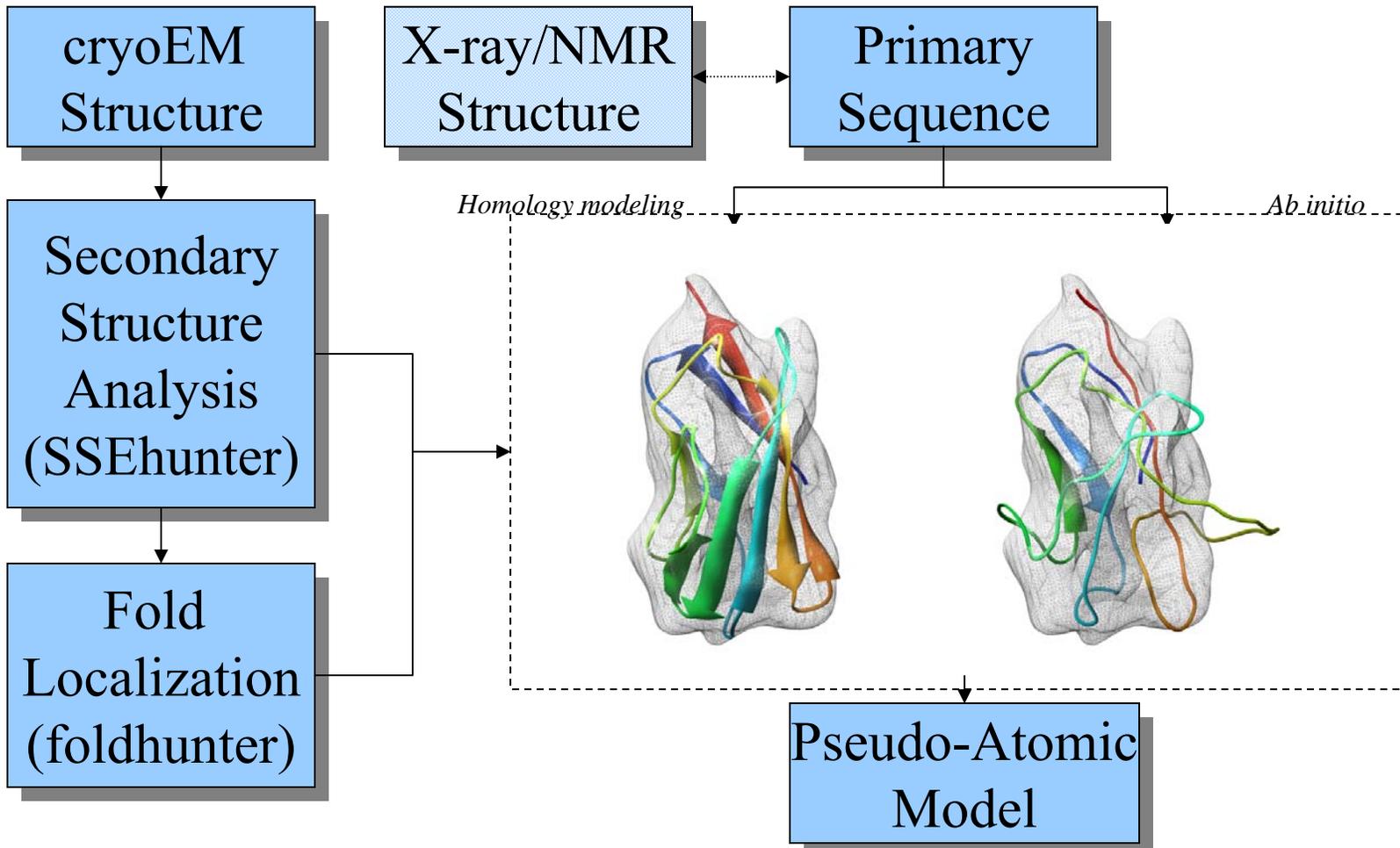
Our approach: top-down and integrate all available knowledge

1. Structural Analysis
 - Identification of SSE w/AIRS (M. Baker)
2. Sequence Analysis
 - Homologue identification
 - Template identification
 - Secondary structure prediction
3. Model Building
 1. SSE assignment
 - Consensus sse assignment
 2. Homology modeling
 - Accurate template models
 3. *ab initio* modeling
 - Domain size limitations

Our Generic Modeling Building Tools



Constrained Modeling



Summary

- Reconstructing CPV to 5.2 Å (including data to 3.5 Å)
- Secondary structure elements and bulky amino-acid side chains are resolved
- Use of an integrative modeling method to build $C\alpha$ models
- Partial atomic model for regions with bulky side chains

Biology: helix as a regulating switch

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